| Test  | Guidelines for Collection  | Collection System  | Optimal<br>Transport                              | Limitations   | Comments   |
|---|--|--|---|---|--|
| Strep A Culture                                   | Depress tongue with tongue depressor. Sample the posterior pharynx and tonsils, avoiding contact with the oral cavity.   | Culture Swab   | Room temp<br>within 1 day                         | Test for the presence of Beta<br>Hemolytic Strep.   | Other Pathogens are covered in a<br>Throat Culture   |
| Clostridium<br>Difficile Molecular                | Collect fresh stool, having the patient pass stool directly into a wide-mouthed container.   | Sterile, leak proof, wide-mouth container  | Within 1 day                                      | Only liquid stools will be tested   | Previously tested patients will not be retested within 7 days of the molecular result.   |
| Culture,<br>Acid Fast and<br>Smear                | Collect appropriate specimen type.  May include tracheal aspirates, bronchial washings, body fluids, stool, CSF, and tissue  Sputum should be 3 complete, consecutive, first morning specimen of at least 5ml.  Urine should be 3 complete, consecutive, first morning specimen. | Sterile, leak proof container  | Within 1 day.  If delay is expected, refrigerate. | Specimens on swabs yield<br>sub-optimal results, and will<br>only be processed if aspirated<br>material or tissue is not<br>available               | Results: Smear reports are available within 24 hours. Final culture reports available at 8 weeks or as completed. Physicians and nursing unit will be notified of <i>ALL</i> positive results immediately. |
| Culture, Acid<br>Fast Blood                       | Collect 10ml of blood in a Sodium Heparin tube.  | Sodium Heparin<br>Tube   | Room temp<br>Within 48<br>hours.                  | Specimen sent to Reference Laboratory for testing.  | Results: 8 weeks or as completed.  |
| Culture, Aerobic /<br>Anaerobic Swabs<br>and Gram | Collect 2 swabs. One Culture Swab One Anaerobic Specimen Collector swab (gray top, glass)  | Culture Swab and<br>Anaerobic Specimen<br>Collector Swab<br>(gray top, glass)  | Room temp<br>Within 1 day                         | Anaerobic cultures will not be done on areas normally colonized with anaerobes, such as vagina or mouth.  | Specimens collected on swabs are least desirable. Collect fluids, aspirates, or tissues when possible.   |
| Culture, Blood                                    | Collect specimen according to Nursing/Phlebotomy Guidelines. Clearly indicate date, time, and site of collection on the vial.  | 2 Bactec Vials: 1 Aerobic (blue cap) 8-10 ml, and 1 Anaerobic (purple cap) 8-10 ml Children: 1 Pediatric (pink top) vial at least 0.5 ml, 1-3 ml optimum | Immediate<br>Room Temp<br>Up to 1 day             | No more than 2 cultures may be drawn per day at no less than 1 hour intervals.  However, 2 cultures maybe drawn simultaneously from separate sites. | All cultures are screened for bacteria. Special requests for yeast or fungi should be indicated in comment or phoned to the micro lab. Blood cultures are held for 5 days.                                 |

| Test  | Guidelines for Collection   | Collection System  | Optimal<br>Transport      | Limitations  | Comments  |
|---|---|--|---------------------------|--|---|
| Culture, Bone   | Aseptically collect sample of bone with minimum attached tissue. Place in sterile container. Clearly indicate site. | Sterile, leak proof container  | Immediate                 | Specimens in their entirety will NOT be accepted for culture. (Ex: an entire toe, skin, tissue and bone together.) | Bones are not appropriate for direct smear or Gram stain.   |
| Culture, Bone<br>Marrow                                 | Collect bone marrow in a plain red top tube.  | Red-top,<br>No additive tube.  | Immediate                 |  |   |
| Culture,<br>Bronchial and<br>Gram Stain                 | Collect specimen during a bronchoscopy procedure in a sterile Lukens tube.  | Sterile Lukens tube  | Immediate                 | Includes BAL, bronchial washings, and brushes  | Bronchial test request form must accompany specimen.  |
| Culture, Cath Tip                                       | Cut off ½" – 1" tip of catheter using sterile technique.  Place in sterile container                                | Sterile container  | Immediate                 | Foley catheter tips are inappropriate for testing. Submit urine instead.   |   |
| Culture, CSF and<br>Gram Stain                          | Collect 1ml or more of CSF in a sterile plastic tube.   | Sterile plastic tube. Sterile Spinal Tap tray available from the sterile processing department | Immediate                 |  | All CSF must be transported directly to the Hematology department for Responsibility tracking.    |
| Culture, Ear and<br>Gram Stain                          | Collect specimen on a culture swab. Indicate right/left, inner/outer.   | Culture swab   | Room temp<br>Within 1 day |  | One specimen is acceptable for bacterial and fungal culture.                                      |
| Culture, Eye and<br>Gram Stain                          | Collect on a mini-tip culture swab. Clearly label site.   | Mini-tip culture swab  | Room temp<br>Within 1 day |  | Eye fluids such as vitreous or aqueous fluid should be ordered as a fluid culture.                |
|   |   |  |                           |  | Scleral rings should be ordered as a Scleral Ring Culture.  |
| Culture, Fluid-<br>Aerobic/Anaerobi<br>c and Gram Stain | Collect fluid in a sterile, plain, NO - ADDITIVE tube. Clearly indicate fluid type.                                 | Sterile, plain, NO-ADDITIVE tube.  | Immediate                 | Swabs should <i>NOT</i> be used in the collection of a fluid.  | All fluids must be transported directly to the Hematology department for Responsibility tracking. |

| Test                                 | Guidelines for Collection   | Collection System   | Optimal<br>Transport  | Limitations   | Comments   |
|--------------------------------------|---|---|---|---|--|
| Culture, Fungus<br>Skin, Hair, Nails | Various specimen types include: Hair clippings, nail clippings / scrapings, skin scrapings. Prior to collection of nail specimens, the surface of the nail should be thoroughly cleaned with 70% alcohol. | A DRY sterile container.                                  | Room temp<br>As soon as<br>possible                             |   | Testing includes a direct exam   |
| Culture, Fungus<br>Screen            | Collect appropriate specimen and inoculate Fungus Media   | Fungus Media,<br>Derma Tube<br>(orange media)             | Room temp As soon as possible                                   |   |  |
| Culture, Fungus<br>Other             | For specimens other than skin, hair, or nails, such as fluids, respiratory specimens, tissues.  | Sterile container   | As soon as possible   | Specimens received on swabs yield sub-optimal results and will only be processed if aspirated material or tissue is not available.                      | Testing includes a direct exam. The direct exam for CSF is an India Ink Prep         |
| Culture, Genital<br>and Gram Stain   | Collect appropriate specimen. Indicate source; cervical, vaginal, penile, urethra, etc.   | Male: mini-tip culture<br>swab<br>Female: culture<br>swab | Room temp<br>Within 1 day                                       |   |  |
| Culture,<br>Legionella               | Collect appropriate specimen in sterile container. Respiratory sources may include; Sputum, Trach, Bronchial Washings.  | Sterile container or<br>Lukens's Tube                     | As soon as possible   |   |  |
| Culture, Scleral<br>Ring             | Collect scleral ring and holding media  | Sterile cup. Capped syringe, WITHOUT needle.              | Immediate   |   |  |
| Culture, Sputum<br>and Gram Stain    | Patient should be instructed to remove dentures, rinse mouth, and gargle with water. The patient should then be instructed to cough deeply and expectorate sputum directly into sterile container.        | Sterile, leak-proof container                             | Room temp,<br>within 2 hours.<br>Refrigerated,<br>within 1 day. | Specimen quality will be evaluated by gram stain.  If it contains oropharyngeal contamination the culture will be canceled and physician/unit notified. | Swabs of sputum will not be processed with the exception of Cystic Fibrosis patients |

| Test  | Guidelines for Collection   | Collection System  | Optimal<br>Transport  | Limitations  | Comments                                     |
|---|---|--|---|--|--|
| Culture, Throat   | Depress tongue with tongue depressor. Sample the posterior pharynx and tonsils, avoiding contact with oral cavity.                                    | Culture Swab   | Room temp<br>Within 1 day                                       |  |  |
| Culture, Tissue<br>Aerobic/Anaerobi<br>c and Gram Stain | Collect surgical or biopsy material in a sterile container. Clearly indicate site/source  | Sterile cup May add sterile saline to prevent specimen from drying out.              | Immediate   | Specimens, in their entirety, will NOT be accepted for culture (Ex: an entire toe, skin tissue and bone together.) |  |
| Culture, Trach/ET<br>and Gram Stain                     | Collect secretions in a sterile Lukens's Tube.  | Sterile Lukens's<br>Tube   | Room temp,<br>within 2 hours.<br>Refrigerated,<br>within 1 day. | Swabs of Trach sites are inappropriate for Trach Culture. Order as Culture, Wound and indicate source.             |  |
| Culture, Urine  | Collect specimen according to Nursing procedures.  Provide patient with clear instructions about cleansing procedures for clean void urine specimens. | Sterile leak-proof<br>container.<br>BD Urine collection<br>device (gray top<br>tube) | Room temp,<br>within 2 hours.<br>BD tube,<br>within 2 days.     |  |  |
| Culture, Vaginal<br>Strep, Group B                      | Collect both vaginal and rectal sources.  | Culture Swab   | Within 2 days   |  |  |
| Culture, Wound<br>and Gram Stain                        | Collect appropriate source.  Label with specific source.  Specify deep or superficial.  | Culture Swab   | Within 1 day  |  |  |
| Culture, Yeast<br>and Smear                             | Collect appropriate specimen.  Various specimen types include; throat, esophageal brush, vaginal, rectal or stool.                                    | Culture Swab<br>Sterile cup  | Within 2 days   |  | Direct exam not performed on urine or stool. |

| Test                                 | Guidelines for Collection   | Collection System   | Optimal<br>Transport   | Limitations  | Comments  |
|--------------------------------------|---|---|--|--|---|
| Enteric<br>Pathogens Panel<br>(NAAT) | Collect liquid or soft stool.  Specimen can be placed in Cary Blair medium if available.  | Sterile leak-proof container. Cary Blair medium.                                      | Room temp,<br>within 1 hour.<br>Refrigerated,<br>within 1 day.<br>Preserved,<br>within 2 days  | Note: Preserved specimens are unacceptable for Toxigenic Clostridium Difficile testing by PCR. | Includes testing for Campylobacter<br>Group, Salmonella Species,<br>Shigella Species, Vibrio Group,<br>Hemorrhagic E coli, Yersinia<br>enterocolitica, Norovirus GI/GII and<br>Rotavirus A. |
| Fecal Leukocytes                     | Collect fresh stool, having patient pass stool directly into widemouthed container.   | Sterile leak-proof container  | Room temp,<br>within 1 hour.<br>Refrigerated,<br>within 1 day.                                 |  |   |
| GC Screen                            | Collect appropriate specimen.  Specimens may include vaginal, rectal, throat. Maybe inoculated directly to media (JEMBEC) available from Microbiology department. | Culture Swab or inoculated JEMBEC media   | Swab – as<br>soon as<br>possible<br>JEMBEC<br>media – as<br>soon as<br>possible                | Timely transport and incubation are critical for the recovery of this organism.                |   |
| GC/<br>Chlamydia by<br>NAAT          | Collect appropriate specimen. Sources may include; endocervical, vaginal, urethral, urine, throat, and rectal.  | Thin Prep Vial APTIMA Unisex or Vaginal Collection Kit                                | Room temp<br>Within 7 days.  | Repeat testing is not indicated within 3 months of treatment.                                  | Test of cure is not indicated per CDC recommendations.  |
|                                      |   | Urine, collect in sterile cup   | Room temp,<br>within 2 hours.<br>Refrigerated,<br>within 1 day.                                |  |   |
| Giardia Antigen<br>Test              | Collect fresh stool, having patient pass stool directly into widemouthed container.  Transport media is available for outpatient collections                      | Sterile leak-proof<br>container<br>OR<br>Para-Pak<br>Pink/Gray Top<br>transport media | Room temp,<br>within 1 hour.<br>Refrigerated,<br>within 1 day.<br>Preserved,<br>within 2 days. |  |   |

| Test  | Guidelines for Collection   | Collection System  | Optimal<br>Transport  | Limitations   | Comments  |
|---|---|--|---|---|---|
| Gram Stain Only                                 | Collect appropriate specimen.   | Culture Swab   | Within 1 day.   |   |   |
| Herpes Molecular<br>(HSV I,<br>HSV II, and VZV) | Vesicular Lesion: Break vesicle and collect fluid and cells from base of lesion.  Non Vesicular Lesion: Collect cells from base of lesion using a swab pre-moistened with saline. | Flocked swab<br>placed in Universal<br>Transport Medium<br>(UTM)                               | Room temp,<br>within 1 day.<br>Refrigerated,<br>within 3 days | NOTE: Prenatal screening specimens are not acceptable for this assay. | Tests for Herpes Simplex Virus Type 1 Herpes Simplex Virus Type 2 and Varicella Zoster Virus. |
| HPV by NAAT                                     | Collect appropriate specimen.   | Thin-Prep Vial   | Room temp<br>Within 7 days                                    |   | HPV Genotyping (16, 18/45) is performed on all positive specimens.                            |
| India Ink Prep                                  | Collect 1 ml or more of CSF in a sterile plastic tube.  | Sterile plastic tube Sterile Spinal Tap tray available from the sterile processing department. | Immediate   |   | India Ink Prep is also included as part of the fungal culture for CSF specimens.              |
| Influenza A/B and<br>RSV by PCR                 | Collect specimen according to Nursing Procedures. Place swab in Universal Transport Media   | Nasopharyngeal<br>mini-tip swab in<br>Universal Transport<br>Media (UTM)                       | As soon as<br>possible, but<br>within 3 days<br>in UTM        |   | All specimens are tested for Influenza A, Influenza B, and Respiratory Syncytial Virus.       |
| KOH Prep  | Collect appropriate specimen in a sterile cup.  Appropriate specimens include skin, hair and nails.   | A DRY sterile container.   | Room temp.<br>As soon as<br>possible.                         |   | This is a direct exam for yeast and fungus.   |

| Test                                       | Guidelines for Collection  | Collection System  | Optimal<br>Transport   | Limitations   | Comments   |
|--|--|--|--|---|--|
| Meningitis<br>Encephalitis<br>Panel (NAAT) | Collect at least 200 microliters of CSF from a lumbar puncture.  | Sterile leak-<br>proof container   | As soon as possible, but room temp for up to 24 hours. Refrigerated, up to 7 days. | CSF obtained via lumbar puncture is the only appropriate specimen for this assay. | Tests for: Escherichia coli K1, Haemophilus influenza, Listeria monocytogenes, Neisseria meningitidis, Strep agalactiae, Strep pneumoniae, Cytomegalovirus, Enterovirus, Herpes simplex virus 1, 2 and 6, Human parechovirus, Varicella zoster virus, Cryptococcus neoformans/gattii |
| Legionella<br>Urinary Antigen              | Collect 1ml or more of urine in a sterile container.   | Sterile leak-proof container   | Room temp,<br>within 2 hours.<br>Refrigerated,<br>within 1 day.                    |   |  |
| MRSA Screen                                | Collect specimen according to Nursing procedures. Source: Nares  | Culture Swab   | Room temp<br>Within 1 day  | Cultures for MRSA only Only Nares will be cultured.                               | MRSA= Methicillin Resistant Staph Aureus   |
| Pinworm<br>Examination                     | Collect a pinworm paddle preparation or perianal region after the patient has been resting at least two hours or in early morning before patient has bathed or defecated | Swube: a pinworm paddle in a sterile plastic tube. Available from the Microbiology department. | Within 1 day   | Do NOT collect on "scotch tape".  Tape preps will NOT be processed.               | Full collection procedures available with <i>Swube</i> collection device.  |

| Test   | Guidelines for Collection  | Collection System  | Optimal<br>Transport  | Limitations  | Comments  |
|--|--|--|---|--|---|
| Respiratory Viral<br>Pathogens by<br>NAAT      | Insert flexible fine-shafted flocked nasopharyngeal swab through nostril into nasopharynx and rotate swab gently a few times.    | Flocked<br>nasopharyngeal<br>swab placed in<br>Universal Transport<br>Medium (UTM) | Room Temp:<br>within 4 hours<br>Refrigerated:<br>within 2 days. |  | RVPM tests for Influenza A,<br>Subtype H1, H1-2009 and<br>Subtype H3, Influenza B,<br>Parainfluenzae 1,2,3,4,<br>Adenovirus, RSV A, RSV B,<br>Human Metapneumovirus,<br>Rhinovirus/Enterovirus,<br>Coronavirus (229E, HKU1, NL63,<br>OC43), SARS-COV-2, Chlamydia<br>pneumoniae and mycoplasma<br>pneumoniae. |
| Scabies  | Collect skin scrapings of lesion according to nursing procedures.  Send glass slide with glycerol and coverslip to lab in a cup. | Glass slide with glycerol and coverslip.   | Within 1 day  | Specimen of choice is a skin biopsy                                    | Itch mites, Sarcoptes scabiei Supplies are available upon request from the Microbiology department.   |
| Staph Screen                                   | Collect specimen according to Nursing procedures. Sources may include nares, axilla, and groin. Primary source: Nares            | Culture Swab   | Room temp<br>Within 1 day                                       | Cultures for Staph aureus-<br>Both sensitive and resistant<br>strains. |   |
| Streptococcus<br>pneumoniae<br>Urinary Antigen | Collect 1 ml or more of urine in a sterile cup.  | Sterile leak-proof container.  | Room temp,<br>within 2 hours.<br>Refrigerated,<br>within 1 day. |  |   |
| Trichomonas by<br>NAAT                         | Collect appropriate specimen. Sources may include; endocervical, vaginal   | ThinPrep Vial APTIMA Unisex or Vaginal Collection Kit.                             | Room temp<br>Within 7 days                                      |  | Not performed on urine or male urethral swabs. These specimens will be sent to a reference laboratory for testing.  |
| VRE Screen                                     | Collect specimen according to Nursing procedures.  | Culture Swab   | Within 1 day  | Source: Peri-rectal swab only  |   |

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